## Siastatin B, a Potent Neuraminidase Inhibitor: The **Total Synthesis and Absolute Configuration**

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Neuraminidase (sialidase, N-acetylneuraminate glycohydrolase EC 3.2.1.18),<sup>1</sup> widely distributed among animal tissues and microorganisms, is involved in various biological functions such as immune response,<sup>2</sup> oncogenesis,<sup>3</sup> metastasis of tumors,<sup>4</sup> sperm penetration,<sup>5</sup> and viral infection,<sup>6</sup> etc.

Siastatin B, a potent inhibitor of neuraminidase, was isolated by Umezawa et al.<sup>7</sup> in 1974 from a Streptomyces culture. Siastatin B inhibits neuraminidases isolated from various microorganisms, animal tissues, and viruses as well as  $\beta$ -glucuronidase and N-acetyl- $\beta$ -D-glucosaminidase. The relative configuration of siastatin B was determined as 2(S/R)-acetamido-3(S/R),4-(R/S)-dihydroxypiperidine-5(R/S)-carboxylic acid by <sup>1</sup>H NMR and X-ray crystallographic studies.<sup>7</sup> However, its absolute configuration was unresolved. We speculated from its biological activity that the absolute configuration of siastatin B should be that shown in 1 by analogy with N-acetylneuraminic acid (2).

Here we wish to report the first total syntheses of siastatin B (1) and its enantiomer (3) based on a chiron strategy. Compound 1 has an unusual structure posessing the continuous -CH-(NHAc)-NH-CH2-CH(COOH)- constituent in a framework. It is distinct from glycohydrolase inhibitors belonging to the sugar analogues having a piperidine ring such as nojirimycin,<sup>8</sup> galactostatin,<sup>9</sup> and their congeners.<sup>10</sup> Our key intermediate for the synthesis of 1 was lactam 6. The synthesis of 6 began with L-ribose which was transformed to 5-azido-5-deoxy-2,3-O-isopropylidene-L-ribonolactone (5),  $[\alpha]^{22}_{D}$  -16.2° (CHCl<sub>3</sub>), by protection of the 2,3-diol, azide formation, and oxidation<sup>11</sup> (Scheme I). Hydrogenation of the amide group of **5** and ring expansion<sup>12</sup> afforded crystalline **6**,  $[\alpha]_{D}^{22}$  -16.4° (CHCl<sub>3</sub>), mp 138-140 °C, in good yield. Stereospecific introduction of the hydroxyl group at C(2) was best achieved by hydride reduction of the protected lactam 7 to 8,  $[\alpha]^{22}_{D}$  +21° (CHCl<sub>3</sub>), mp 164 °C, and Swern oxidation<sup>13</sup> to give aminal 9,  $[\alpha]^{22}_{D}$  +11° (CHCl<sub>3</sub>), mp 106-107

(1) Gottshalk, K. The Chemistry and Biology of Sialic Acid and Related Substances; Cambridge University Press: London, 1960; pp 75-87. Roseman, S. Chem. Phys. Lipids 1970, 5, 270. (2) Simons, R. L.; Rios, A. J. Immunol. 1973, 111, 1820. Rott, R.; Becht,

H.; Orlich, M. J. Gen. Virol. 1974, 22, 35.

(3) Bekesi, J. G.; St-Arneault, G.; Holland, J. F. Cancer Res. 1971, 31,

 Viscos, S. G., Strundadt, G., Honald, S. F. Cunter Res. 1971, 31, 2130. Bernacki, R. J. Science (Washington, D.C.) 1977, 195, 577.
 (4) Gasic, G.; Gasic, T. Proc. Nat. Acad. Sci. U.S.A. 1962, 48, 1172.
 Yogeeswaran, G.; Salk, P. L. Science (Washington, D.C.) 1981, 212, 1514.
 Dennis, J.; Waller, C.; Timpl, R.; Schirrmacher, V. Nature (London) 1982, 300, 274.
 Fogel, M.; Altevogt, P.; Schirrmacher, V. J. Exp. Med. 1983, 157, 371 371

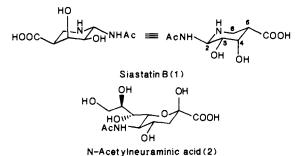
(5) Soupart, P. S.; Clewe, T. H. Fert. Steril. 1965, 16, 677.
(6) Ackermann, W. W.; Ishida, N.; Massab, H. F. J. Exptl. Med. 1955, 102, 545. Haff, R. F.; Stewart, R. C. J. Immunol. 1965, 94, 842. Lipkind, M. A.; Tsvetkova, I. V. Arch. Ges. Virusforsch. 1970, 29, 370. Huang, R. T. C.; Rott, R.; Wahn, K.; Klenk, H. D.; Kohama, T. Virology 1980, 107, 313.
(7) Umogene H. Acuesia, T. V. Swingene, T. Masikara, M. S. Marada, S. S. Ma

 C.; Rott, R.; Wann, K.; Klenk, H. D.; Kohama, I. Virology 1980, 107, 315.
 (7) Umezawa, H.; Aoyagi, T.; Komiyama, T.; Morishima, H.; Hamada, M.; Takeuchi, T. J. Antibiot. 1974, 27, 963.
 (8) Inouye, S.; Tsuruoka, T.; Niida, T. J. Antibiot. 1966, Ser. A19, 288.
 (9) Miyake, Y.; Ebata, M. J. Antibiot. 1987, 40, 122.
 (10) Inouye, S.; Tsuruoka, T.; Ito, T.; Niida, T. Tetrahedron 1968, 23, 2125. Schmidt, D. D.; Frommer, W.; Müller, L.; Truscheit, E. Naturwis-senschaften 1979, 66, 584. Fleet, G. W. J.; Smith, W.; Evans, S. V.; Fellows, L. E. J. Chem. Soc., Chem. Commun. 1984, 1240. Fellows, L. E.; Bell, E. A.: Lyon D. G.; Pilkiewicz, F. Nakanishi, K. J. Chem. Soc. Chem. Commun. .; Lynn, D. G.; Pilkiewicz, F.; Nakanishi, K. J. Chem. Soc., Chem. Commun. 1979, 977

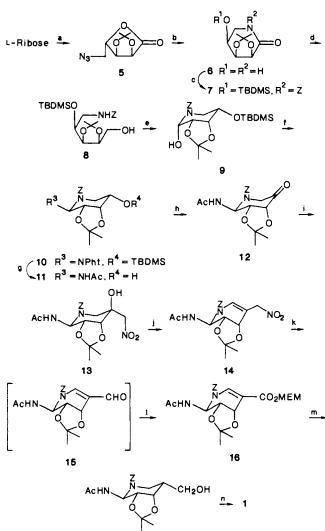
 (11) Hanessian, S. J. Org. Chem. 1969, 34, 675.
 (12) For a review, see: Paulsen, H.; Todt, K. In Advances in Carbohydrate Chemistry: Cyclic Monosaccharides Having Nitrogen or Sulfur in the Ring; Wolfrom, M. L., Tipson, R. T., Eds.; Academic Press: New York, 1968; Vol.

23, pp 115-232. (13) Mancuso, A. J.; Huang, S.-L.; Swern, D. J. Org. Chem. 1978, 43, 2480.

Chart I



Scheme I<sup>a</sup>



17

<sup>a</sup>(a) p-TsOH, Me<sub>2</sub>CO; CH<sub>3</sub>SO<sub>2</sub>Cl, C<sub>6</sub>H<sub>5</sub>N; NaN<sub>3</sub>, DMSO; CrO<sub>3</sub>/ C<sub>6</sub>H<sub>5</sub>N, CH<sub>2</sub>Cl<sub>2</sub>, 89%; (b) H<sub>2</sub>, Raney Ni, MeOH, 88%; (c) t-BuMe<sub>2</sub>SiCl, imidazole, DMF; PhCH<sub>2</sub>OCOCl, NaH, DMF, 99%; (d) NaBH<sub>4</sub>, EtOH, 70%; (e) (COCl)<sub>2</sub>, DMSO, Et<sub>3</sub>N, CH<sub>2</sub>Cl<sub>2</sub>, 88%; (f) phthalimide, Ph<sub>3</sub>P, DEAD, DMF, 100%; (g) NH<sub>2</sub>NH<sub>2</sub>, MeOH; Ac<sub>2</sub>O,  $C_6H_5N$ ; *n*-Bu<sub>4</sub>NF, THF, 100%; (h) RuO<sub>4</sub>,  $CH_2Cl_2/CCl_4$ , 99%; (i) CH<sub>3</sub>NO<sub>2</sub>, NaH, DME, 100%; (j) p-TsOH, Ac<sub>2</sub>O; K<sub>2</sub>CO<sub>3</sub>, C<sub>6</sub>H<sub>6</sub>, 100%; (k) C<sub>6</sub>H<sub>5</sub>N, 38 °C, 80%; (l) CH<sub>3</sub>CH=C(CH<sub>3</sub>)<sub>2</sub>/t-BuOH, NaOCl<sub>2</sub>-NaH<sub>2</sub>O<sub>4</sub>/H<sub>2</sub>O; MEMCI,  $(i-Pr)_2$ NEt, CH<sub>2</sub>Cl<sub>2</sub>, 55%; (m) NaBH<sub>4</sub>, 1:10 CF<sub>3</sub>CH<sub>2</sub>OH/THF, 75%; (n) PDC, DMF; H<sub>2</sub>, 5% Pd/C, MeOH; 1 M aqueous HCl, then Dowex 50W-X4 (H<sup>+</sup> form) eluted with NH<sub>4</sub>-OH, 66%.

°C. The <sup>1</sup>H NMR spectrum of 9 shows a proton of C(2) at  $\delta$ 5.55 (a singlet with a small coupling, J < 2 Hz), clearly indicative of an equatorial hydrogen. Strikingly a single stereoisomer

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controlled by an anomeric effect<sup>14</sup> results from this oxidation, whereas oxidation with CrO<sub>3</sub> in pyridine gives a 2:1 mixture of 9 and its epimer at C(2). Displacement of the axial hydroxyl group to the equatorial amino group proved troublesome until we discovered that the Mitsunobu reaction<sup>15</sup> (PPh<sub>3</sub>, diethyl azodicarboxylate, phthalimide) in N,N-dimethylformamide gave the desired product 10,  $[\alpha]^{22}_{D}$  +26.4° (CHCl<sub>3</sub>), mp 109 °C (dec), quantitatively. The stereochemistry at C(2) was established by its <sup>1</sup> H NMR spectrum which shows a proton of C(2) at  $\delta$  5.91 (d, J = 5 Hz), clearly indicative of an axial hydrogen. Replacement of the amino substituent and removal of the tert-butyldimethylsilyl group to 11,  $[\alpha]^{22}_{D}$  -20.4° (CHCl<sub>3</sub>), and oxidation to 12,  $[\alpha]^{22}_{D}$  -56° (CHCl<sub>3</sub>), were unexceptional.

Condensation of 12 with nitromethane was found to proceed smoothly to give 13 as a single stereoisomer,  $[\alpha]^{22}_{D} + 26^{\circ}$  (CHCl<sub>3</sub>), quantitatively. The stereochemistry at C(5) is tentatively assigned as 13. Acetylation of 13 followed by base-catalyzed elimination of the acetoxy group afforded 14,  $[\alpha]^{22}_{D}$  +69° (CHCl<sub>3</sub>), in a good yield. The structure of 14 was determined by its <sup>1</sup> H NMR spectrum which shows the methylene protons of the nitromethyl group at  $\delta$  4.78 and 5.16 (ABq, J = 15 Hz), a proton of C(4) at  $\delta$  4.71 (d, J = 6 Hz), and a proton of C(6) at  $\delta$  7.06 (S). Transformation of 14 to carboxylate 16,  $[\alpha]^{22}_{D}$  -4.1° (MeOH), mp 194–198 °C (dec), was achieved via  $\alpha,\beta$ -unsaturated aldehyde 15 produced by simply warming 14 in pyridine. Compound 16 was also successfully produced from 13 by the stepwise successive sequences without isolation.

A problem arose, however, as catalytic reduction<sup>16</sup> of 16 accompanied by elimination of the hydroxyl group at C(4) and hydride reduction<sup>17</sup> of the double bond proceeded unfavorably and without chemoselectivity. To circumvent this problem, 16 was stereoselectively hydrogenated to  $\alpha,\beta$ -saturated hydroxymethyl compound 17 (NaBH<sub>4</sub>, 1:10 CF<sub>3</sub>CH<sub>2</sub>OH/THF),  $[\alpha]^{22}_{D}$  +7.0° (CHCl<sub>3</sub>). The <sup>1</sup>H NMR spectrum of 17 shows protons at  $\delta$  3.25 (t, J = 13 Hz, H-6 ax), 3.59 (dd, J = 5 and 13 Hz, H-6 eq), and4.54 (dd, J = 3 and 8 Hz, H-4), clearly indicative of an axial hydrogen at C(5).

The remaining steps of the synthesis are rather straightforward. The carboxylic acid formed upon oxidation of 17 was converted by removal of protecting groups to crystalline 1,  $[\alpha]^{22}_{D} + 56^{\circ}$  (H<sub>2</sub>O) (lit.<sup>7</sup> 57.2° (H<sub>2</sub>O)), mp 135–136 °C (dec) (lit.<sup>7</sup> 137 °C), in 10.3% overall yield from L-ribose. Its spectral properties (IR, <sup>1</sup>H NMR, <sup>13</sup>C NMR, mass spectrum) were superimposable with those of the natural specimen.

The enantiomer 3 was also synthesized from D-ribonolactam by the same method used in the synthesis of 1. Compound 3 was identical in all respects with the synthetic and the natural 1 except for the sign of the specific rotation.

Thus, the absolute configuration of siastatin B has been elucidated as the (2R, 3R, 4S, 5S)-isomer 1.

The synthetic 1 shows the same inhibitory effects as the natural one against neuraminidases prepared from Cl. perfringens, Streptomyces, rat mammary gland, rat mammary liver, chorioallantoic membrane (IC<sub>50</sub> = 3, 10, 110, 170 and 55  $\mu$ g/0.5 mL, respectively),  $\beta$ -glucuronidase (IC<sub>50</sub> = 4  $\mu$ g/0.5 mL), and Nacetyl- $\beta$ -D-glucosaminidase (IC<sub>50</sub> = 18  $\mu$ g/0.5 mL).<sup>7</sup> In contrast, compound 3 shows only weak activities against neuraminidases mentioned above (IC<sub>50</sub>: more than 100  $\mu$ g/0.5 mL), whereas 3 demonstrates activity against  $\beta$ -glucuronidase (IC<sub>50</sub> = 25  $\mu$ g/0.5 mL).

## **Observation and Substituent Control of** Medium-Dependent Hot-Molecule Reactions in Low-Temperature Matrices

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Although reaction from vibrationally excited products is common in the gas phase, hot-molecule chemistry in condensed phases is rare.<sup>2</sup> Vibrational energy transfer to the medium is generally faster than chemically activated reaction. There are, however, a handful of cases where hot-molecule reactions of ground-state products of thermal<sup>3-5</sup> or photochemical<sup>6-8</sup> processes in various media have been proposed to explain unexpected products. Reaction from upper vibrational levels in electronic excited states has also been suggested in several instances<sup>9-11</sup> to rationalize wavelength dependence in condensed phase photochemistry.

There is spectroscopic evidence that vibrational relaxation in inert-gas matrices may be slow relative to solution. For example, Bondybey<sup>12</sup> and Rentzepis<sup>13</sup> have observed relatively long-lived hot bands (ca. 300 ps) in the fluorescence spectra of various aromatics in rare-gas matrices at 4 K. The higher energy vibrational emissions are absent in hydrocarbon matrices<sup>14</sup> or when the substrates are methylated.<sup>12b</sup> The surprising lifetimes have been attributed to mismatch of the vibrations of the aromatics and the low-energy lattice modes of the matrices, leading to poor energy transfer.<sup>12,13</sup> IR induced conformational interconversions in matrices are also well known.5 The possibility of slow vibrational relaxation of photoproducts raises concerns for the observation of highly reactive intermediates with matrix-isolation techniques. We now wish to report chemical evidence, based on media and substituent effects, for the formation of vibrationally hot photoproducts in low-temperature matrices and their subsequent ground-state reactions.

We have reported that irradiation of 1 in toluene- $d_8$  at 77 K gives primarily 7-norbornadienone (2), characterized by <sup>1</sup>H NMR at  $-78^{\circ}$  C.<sup>15,16</sup> A minor product, tentatively assigned as 3, was also observed. On warming to ca. -60 °C, 2 cleanly gave benzene (and CO), with  $\Delta G^* = 15$  kcal/mol. Side product 3 polymerized

- (2) Bondybey, V. E. Ann. Rev. Phys. Chem. 1984, 35, 591 and references cited therein.
- (3) Farneth, W. E.; D'Amore, M. B.; Brauman, J. I. J. Am. Chem. Soc. 1976, 98, 5546.
- (4) Rohman, M.; Shevlin, P. Tetrahedron Lett. 1985, 26, 2959. (5) Frei, H.; Pimentel, G. C. Ann. Rev. Phys. Chem. 1985, 36, 491 and references therein
- (6) Teki, Y.; Takui, R.; Itoh, K.; Iwamura, H.; Kobaayashi, K. J. Am. Chem. Soc. 1986, 108, 2147.
- (7) Jain, R.; McElwee-White, L.; Dougherty, D. A. J. Am. Chem. Soc. 1988, 110, 552.

(8) Vibrational lifetimes in diatomics can be extremely long in condensed phases,<sup>2</sup> and this can play a role in their chemistry. See, for example: Smith, D. E.; Harris, C. B. J. Chem. Phys. 1987, 87, 2709.
(9) (a) Castellan, A.; Kolc, J.; Michl, J. J. Am. Chem. Soc. 1978, 100, 6687.
(b) Souto, M. A.; Kolc, J.: Michl, J. J. Am. Chem. Soc. 1978, 100,

6692.

(10) Mauring, L. E.; Peters, K. S. J. Am. Chem. Soc. 1984, 106, 8077. (11) Barbara, P. F.; Brus, L. E.; Rentzepis, P. M. J. Am. Chem. Soc. 1980, *102*, 5631.

(12) (a) Huppert, D.; Bondybey, V. E.; Rentzepis, P. M. J. Phys. Chem. 1985, 89, 5811. (b) Bondybey, V. E.; Haddon, R. C.; Rentzepis, P. M. J. Am. Chem. Soc. 1984, 106, 5969. (c) Rentzepis, P. M.; Bondybey, V. E. J. Chem. Phys. 1984, 80, 4727

Phys. 1984, 80, 4127.
(13) (a) Hopkins, J. B.; Rentzepis, P. M. Chem. Phys. Lett. 1985, 117, 414. (b) Hopkins, J. B.; Rentzepis, P. M. Chem. Phys. Lett. 1987, 142, 503.
(c) Huppert, D.; Rentzepis, P. M. J. Chem. Phys. 1987, 86, 603.
(14) Gillispie, G. D. J. Chem. Phys. 1985, 85, 4825.
(15) LeBlanc, B. F.; Sheridan, R. S. J. Am. Chem. Soc. 1985, 107, 4554.
(16) For an alternate synthesis and investigation of 7-norbornadienone, see:
Birney, D. M.; Berson, J. A. J. Am. Chem. Soc. 1985, 107, 4553. Birney, D.

M.; Berson, J. A. Tetrahedron 1986, 43, 1561.

0002-7863/88/1510-7250\$01.50/0 © 1988 American Chemical Society

<sup>(14)</sup> Lemieux, R. U.; Kulling, R. K.; Bernstein, H. J.; Schneider, W. G. J. Am. Chem. Soc. 1958, 80, 6098.
(15) For a review, see: Mitsunobu, O. Synthesis 1981, 1.
(16) Hanessian, S.; Lavallee, P. Can. J. Chem. 1977, 55, 562. Anderson,

R. C.; Fraser-Reid, B. Tetrahedron Lett. 1978, 3233. Hanessian, S.; Ranc-

K. C., Fraser-Reid, B. *Tetrahearon Lett.* 1978, 3253. Fanessian, S.; Kancourt, G.; Guindon, Y. Can. J. Chem. 1978, 56, 1843. Corey, E. J.; Goto, G. *Tetrahedron Lett.* 1980, 3463.
 (17) Kadin, S. B. J. Org. Chem. 1966, 31, 620. Satoh, T.; Nanba, K.; Suzuki, S. Chem. Pharm. Bull. 1971, 19, 817. Ganem, B.; Fortunato, J. M. J. Org. Chem. 1975, 40, 2846. Hutchins, R. O.; Rotstein, D.; Natale, N.; Fanelli, J. J. Org. Chem. 1976, 41, 3328. Kumar, Y.; Florvall, L. Synth. Commun. 1983, 13, 489. Setoi, H.; Takeno, H.; Hashimoto, M. J. Org. Chem. 1985, 50, 3948.

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